

Syntheses and Biological Evaluation of Irciniastatin A and the C1–C2 Alkyne Analogue

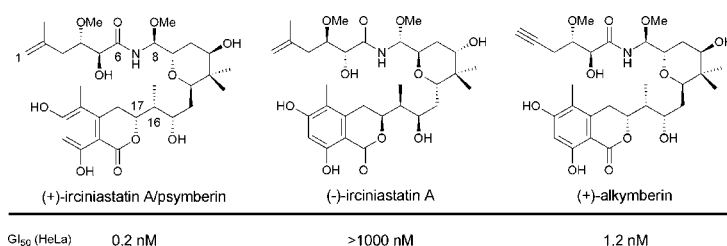
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ABSTRACT



Syntheses of both natural (+)- and unnatural (-)-irciniastatin A (aka psymberin) as well as a C1–C2 alkyne analogue of (+)-irciniastatin A have been achieved. The key features of the syntheses include a highly regioselective epoxide-opening reaction and a late-stage assembly of C1–C6, C8–C16, and C17–C25 fragments. (+)-Alkymberin retained a high level of cytotoxicity, whereas (-)-irciniastatin A showed almost no activity. These results suggest that (+)-alkymberin could be a useful enantio-differential probe for mode-of-action study.

In 2004, (+)-irciniastatin A (**1**)¹ and psymberin,² new pederin-type natural products, were isolated by the Pettit group from marine sponge *Ircinia ramosa* and by the Crews group from marine sponge *Psammocinia* sp. (Figure 1). In addition to (+)-irciniastatin A (**1**), the Pettit group also isolated the C11 ketone analogue, named (-)-irciniastatin B. (+)-Irciniastatin A (**1**) has been shown to exhibit extremely potent and selective cytotoxicity against certain human cancer cell lines.^{1,2}

The promising therapeutic potential coupled with the limited availability of these natural products has attracted significant attention from the synthetic community. In 2005, the first total synthesis of (+)-psymberin was achieved by

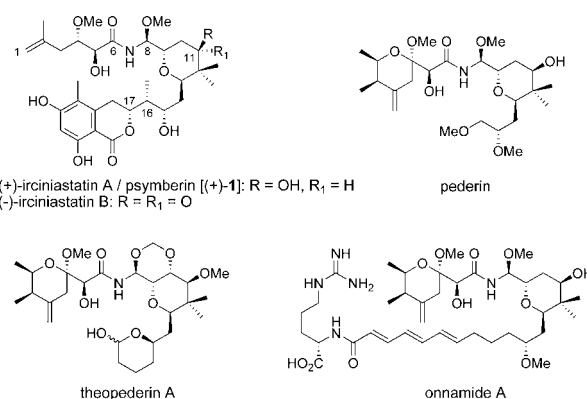


Figure 1. Pederin-type natural products.

the De Brabander group, who demonstrated that (+)-irciniastatin A and (+)-psymberin are identical, as repre-

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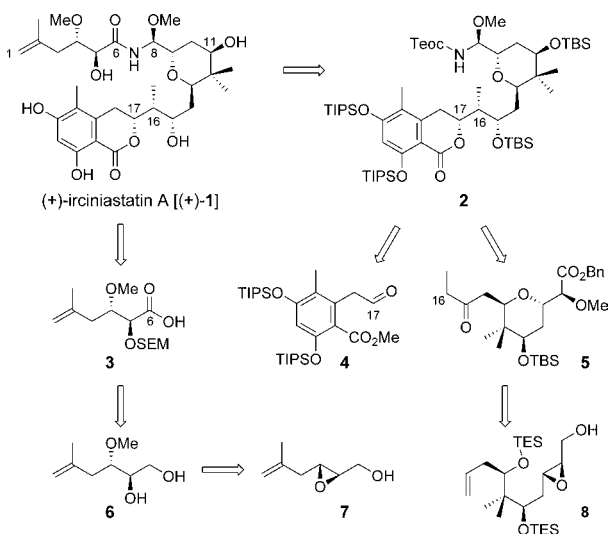
(2) Cichewicz, R. H.; Valeriote, F. A.; Crews, P. *Org. Lett.* **2004**, *6*, 1951.

sented by (+)-**1**.³ To date, several total^{4–6} and formal⁷ syntheses as well as SAR studies^{8,9} have been reported. Among them, the Schering-Plough group has reported that the substituents at C4 and C5 are important for the cytotoxicity, and that the C1–C2 double bond is not essential for activity.⁹ Also, preliminary biological studies using natural and synthetic samples have suggested that (+)-irciniastatin A (**1**) might have a different mode-of-action from that of other pederin family members (Figure 1).^{2,8}

Intrigued by these results, we, several years ago, started a synthetic program that would enable acquisition of possible isomers and analogues. As a part of this program, we herein describe the syntheses and biological evaluation of both enantiomers of irciniastatin A (**1**), and (+)-“alkymberin”, a C1–C2 alkyne analogue of natural (+)-**1**.

Retrosynthetically, (+)-irciniastatin A (**1**) was divided into the C1–C6 acyclic side chain **3** and protected hemiaminal **2**, which in turn disconnected in a retro-aldol fashion into C17–C25 aldehyde fragment **4** and C8–C16 tetrahydropyran fragment **5** (Scheme 1). It should be noted that intermediates **3**

Scheme 1. Retrosynthetic Analysis of (+)-Irciniastatin A (**1**)



and **5** could be derived from epoxy alcohols **7** and **8**, respectively. To synthesize not only fragments **3** and **5** but also their isomers for SAR study, we planned to utilize a Sharpless asymmetric epoxidation (SAE) chemistry¹⁰ as a key reaction. For example, enantiomers and diastereomers of **6** could be

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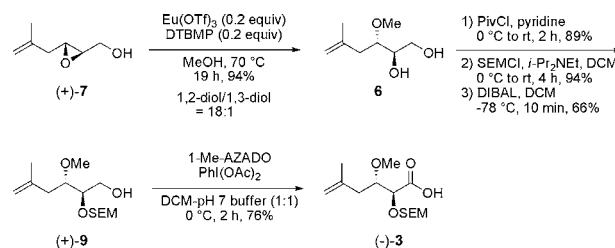
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synthesized by the regioselective ring-opening of those of epoxy alcohol **7**, which could be easily obtained using an SAE strategy.

The synthesis of **3** commenced with regioselective ring-opening of known epoxy alcohol (+)-**7**¹¹ with MeOH (Scheme 2). Initially, we tried Sharpless condition using Ti(OiPr)₄ as

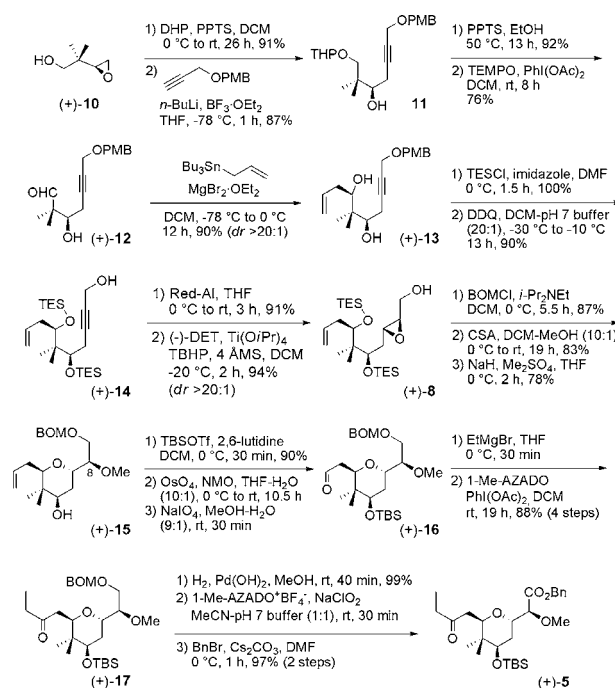
Scheme 2. Synthesis of the Acyclic Side Chain (–)-**3**



Lewis acid,¹² but the yield and selectivity were unsatisfactory (54%, 1,2-diol:1,3-diol = 3:1). To improve this situation, we screened various Lewis acids (BF₃·OEt₂, MgBr₂ etc.) and finally found that Eu(OTf)₃ gave the desired 1,2-diol **6**, which is inseparable from the corresponding 1,3-diol, in high yield and selectivity (>20:1). The loading of Eu(OTf)₃ could be reduced to a catalytic amount when it was used with 0.2 equiv of 2,6-di-*tert*-butyl-4-methylpyridine (DTBMP), giving comparable selectivity (18:1).¹³ The obtained 1,2-diol **6** was converted via the usual three steps to the primary alcohol (+)-**9**, which was oxidized using 1-Me-AZADO¹⁴ to furnish carboxylic acid (–)-**3**.

The synthesis of **5** began with a known epoxy alcohol (+)-**10**,¹⁵ which was derived from commercially available (–)-pantolactone (Scheme 3). After protection of the primary

Scheme 3. Synthesis of C8–C16 Fragment (+)-**5**



hydroxyl group in (+)-**10**, the epoxide was opened regioselectively with lithium acetylide and $\text{BF}_3 \cdot \text{OEt}_2$ ¹⁶ to give alcohol **11**. Deprotection and subsequent TEMPO oxidation afforded aldehyde (+)-**12**.

We then focused on the diastereoselective allylation of (+)-**12**. After extensive experimentation, we found that the reaction using allyltributylstannane and MgBr_2 proceeded in a highly diastereoselective manner to give diol (+)-**13**.¹⁷ Protecting group manipulation and *trans*-reduction of the alkyne moiety provided the corresponding allyl alcohol, which was then subjected to SAE to give (+)-**8** in 94% yield and >20:1 diastereoselectivity. After protection of the primary hydroxyl group, treatment of the epoxide with CSA in $\text{MeOH}-\text{CH}_2\text{Cl}_2$ effected the deprotection of two TES groups and formation of the desired tetrahydropyran core.

Selective methylation of the C8-hydroxyl group was accomplished using Me_2SO_4 to give methyl ether (+)-**15** in 78% yield. Silylation of the remaining secondary alcohol in (+)-**15** followed by oxidative cleavage of the terminal olefin (OsO_4 , NMO; NaIO_4) provided the corresponding aldehyde **16**. Treatment of crude **16** with ethylmagnesium bromide followed by oxidation of the resultant secondary alcohol using 1-Me-AZADO gave ketone (+)-**17** in 88% for 4 steps. Finally, cleavage of BOM ether, oxidation of the resultant alcohol using 1-Me-AZADO⁺ $\text{BF}_4^-/\text{NaClO}_2$,¹⁸ and protection provided benzyl ester (+)-**5** in high yield.

Aldehyde **4** was prepared based on De Brabander's protocol.³ The union of **4** with (+)-**5** was achieved by mixing the *Z*-boron enolate of (+)-**5** with aldehyde **4** at -78°C to give the aldol product (+)-**18** in a highly diastereoselective manner (Scheme 4).¹⁹ Reduction of (+)-**18** with NaBH_4 in

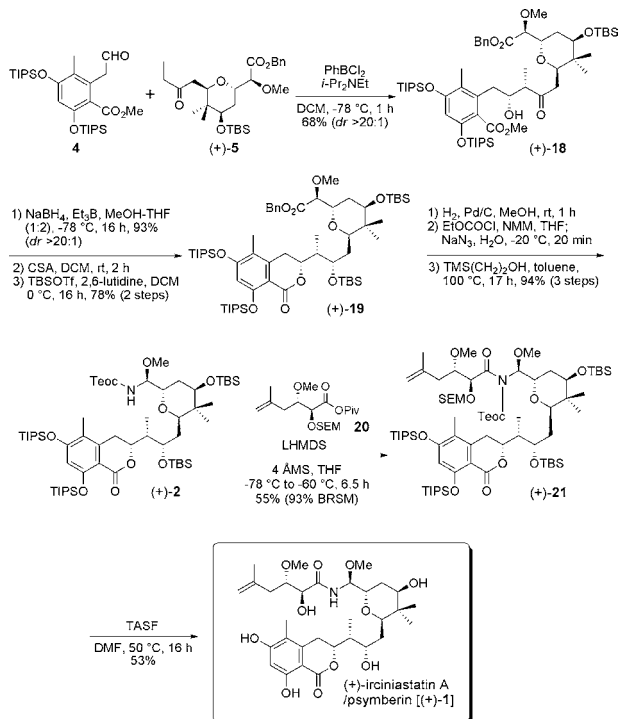
the presence of Et_3B in MeOH provided the 1,3-*syn* diol (dr >20:1),²⁰ which was converted to lactone (+)-**19** in 78% yield in 2 steps. Hydrogenolysis of benzyl ester followed by a Curtius sequence using 2-(trimethylsilyl)ethanol as a nucleophile gave Teoc-protected hemiaminal (+)-**2** in high yield.^{21,22}

Coupling of (+)-**2** with the acyclic side chain fragment proved to be a difficult task. Initially, we examined the coupling reaction of (+)-**2** with several derivatives of the carboxylic acid (–)-**3**, which never yielded the desired product. After intensive effort, we realized that Teoc-protected hemiaminal (+)-**2** and pivalate **20** were most suitable for this coupling reaction.⁵ Finally, global deprotection using TASF provided (+)-irciniastatin A (**1**).

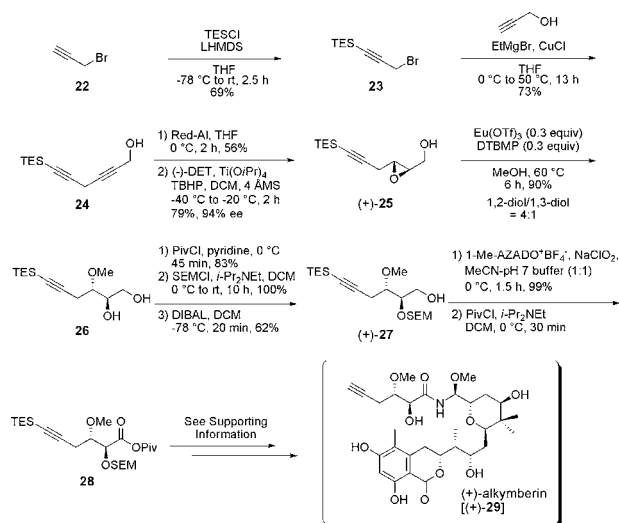
With a highly convergent and flexible route to access (+)-irciniastatin A (**1**) in hand, we then synthesized “alkymberin” (**29**), which bears an alkyne moiety at the C1–C2 position. As described, the C1–C2 olefin moiety has been reported to be unnecessary for cytotoxicity,⁹ and the alkyne part is expected to be a useful handle for introducing several reporter tags using click chemistry.²³ Moreover, we decided to synthesize (–)-irciniastatin A (**1**) to examine whether irciniastatin A acts as a “ligand” or “chemical reagent” in cells. For example, the acyl aminal at C8 could be a good electrophilic reagent (i.e., acylimine) when the methoxy group at C8 was eliminated.

For the synthesis of (+)-alkymberin, we prepared alkyne side chain **28** based on the synthetic route previously established (Scheme 5). In the course of the synthesis, we

Scheme 4. Total Synthesis of (+)-Irciniastatin A (1**)**



Scheme 5. Synthesis of (+)-Alkymberin [(+)-29**]**

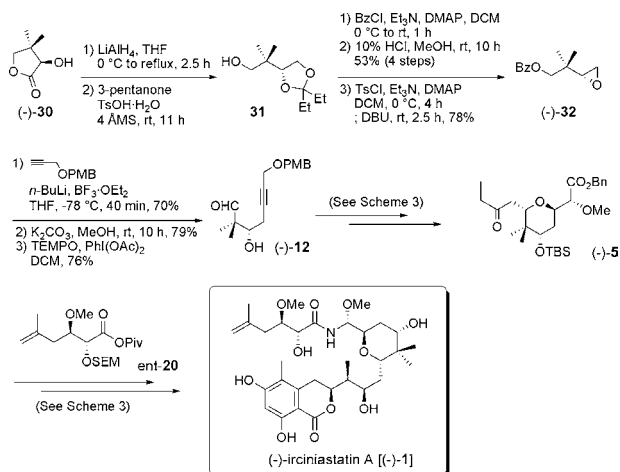


found that the 1-Me-AZADO⁺ $\text{BF}_4^-/\text{NaClO}_2$ system was more effective for the oxidation of alcohol (+)-**27**. The resultant carboxylic acid was activated as mixed anhydride **28**, which was successfully coupled with (+)-**2** to furnish (+)-alkymberin [(+)-**29**].

To synthesize (–)-irciniastatin A (**1**), each enantiomer of the C1–C6 acyclic side chain [i.e., *ent*-**20**] and the C8–C16

tetrahydropyran fragment [(-)-5] is needed. The C1–C6 acyclic side chain *ent*-20 was prepared using (+)-DET by SAE in the established route in Scheme 2. The C8–C16 tetrahydropyran fragment (-)-5 was synthesized from aldehyde (-)-12, which was prepared in 8 steps from (-)-pantolactone (30) (Scheme 6).

Scheme 6. Synthesis of (-)-Irciniastatin A (1)



Synthetic (+)- and (-)-irciniastatin A (1) and (+)-alkymberin (29) were evaluated for their cytotoxicity against HeLa cells. As expected, (+)-alkymberin (29) retained a high level of cytotoxic activity [GI_{50} value of 1.2 nM for (+)-1, 0.2 nM for (+)-29]. In contrast, (-)-irciniastatin A (1) showed almost no cytotoxic activity ($GI_{50} > 1000$ nM). These results indicated that an enantio-differential recognition event occurs between (+)-irciniastatin A (1) and its cellular target;

as such, (+)-alkymberin [(+)-29] is a good candidate for an enantio-differential probe²⁴ for mode-of-action study.¹⁹

In summary, we have accomplished syntheses of (+)- and (-)-irciniastatin A (1), as well as (+)-alkymberin (29), via a convergent synthetic route. Biological evaluation of these compounds suggested that (+)-alkymberin (29) can be a good enantio-differential probe for analyzing mode-of-action of (+)-irciniastatin A (1). Further studies on both SAR and the mode-of-action of irciniastatins are now in progress, and results will be reported in the near future.

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Supporting Information Available: Experimental procedures, characterization data, and copy of NMR spectra. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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